

# Package ‘isocalcR’

July 31, 2021

**Type** Package

**Title** Isotope Calculations in R

**Version** 0.0.2

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**Description** Perform common calculations based on published stable isotope theory, such as calculating carbon isotope discrimination and intrinsic water use efficiency from wood or leaf carbon isotope composition. See Farquhar, O’Leary, and Berry (1982) <[doi:10.1071/PP9820121](https://doi.org/10.1071/PP9820121)>.

**License** GPL-3

**URL** <https://github.com/justinmathias/isocalcR>

**BugReports** <https://github.com/justinmathias/isocalcR/issues>

**Depends** R (>= 4.0.0)

**Imports** dplyr (>= 1.0.6)

**Encoding** UTF-8

**Language** en-US

**LazyData** true

**Suggests** rmarkdown, knitr, testthat (>= 3.0.0)

**VignetteBuilder** knitr

**Config/testthat/edition** 3

**RoxygenNote** 7.1.1

**NeedsCompilation** no

**Repository** CRAN

**Date/Publication** 2021-07-31 11:50:05 UTC

## R topics documented:

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CO2data	<i>CO2data</i>
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### Description

Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. Data are from Belmecheri, Lavergne, 2020, Dendrochronologia. Updated based on their methodology beyond C.E. 2019.

### Usage

```
data(CO2data)
```

### Format

A data frame with 2020 rows and 3 variables:

**yr** Year of CO<sub>2</sub> and d13CO<sub>2</sub> measurement

**Ca** Atmospheric CO<sub>2</sub> concentration, in ppm

**d13C.atm** Atmospheric d13CO<sub>2</sub>, in per mille, ‰

### Source

<https://www.sciencedirect.com/science/article/abs/pii/S1125786520300874>

### References

Belmecheri, S. & Lavergne, A. (2020). Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. Dendrochronologia, 63, 125748.

### Examples

```
data(CO2data)
head(CO2data)
```

---

d13C.to.Ci

*d13C.to.Ci*


---

**Description**

Calculates leaf intercellular CO<sub>2</sub> concentration given plant tissue d13C signature.

**Usage**

d13C.to.Ci(d13C, year, elevation, temp, frac = 0)

**Arguments**

d13C	Measured plant tissue carbon isotope signature, per mille (‰)
year	Year to which the sample corresponds
elevation	Elevation (m.a.s.l.) of the sample, necessary to account for photorespiration processes
temp	Leaf temperature (°C)
frac	Post-photosynthetic fractionation factor, defaults to 0 assuming leaf material, user should supply reasonable value if from wood (generally -1.9 - -2.1)

**Value**

The concentration of leaf intercellular CO<sub>2</sub> (Ci) in parts per million (ppm)

**References**

- Badeck, F.-W., Tcherkez, G., Nogués, S., Piel, C. & Ghashghaie, J. (2005). Post-photosynthetic fractionation of stable carbon isotopes between plant organs—a widespread phenomenon. *Rapid Commun. Mass Spectrom.*, 19, 1381–1391.
- Belmecheri, S. & Lavergne, A. (2020). Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. *Dendrochronologia*, 63, 125748.
- Bernacchi, C.J., Singaas, E.L., Pimentel, C., Portis Jr, A.R. & Long, S.P. (2001). Improved temperature response functions for models of Rubisco-limited photosynthesis. *Plant, Cell Environ.*, 24, 253–259.
- Craig, H. (1953). The geochemistry of the stable carbon isotopes. *Geochim. Cosmochim. Acta*, 3, 53–92.
- Davies, J.A. & Allen, C.D. (1973). Equilibrium, Potential and Actual Evaporation from Cropped Surfaces in Southern Ontario. *J. Appl. Meteorol.*, 12, 649–657.
- Farquhar, G., O’Leary, M. & Berry, J. (1982). On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.*, 9, 121–137.

Frank, D.C., Poulter, B., Saurer, M., Esper, J., Huntingford, C., Helle, G., et al. (2015). Water-use efficiency and transpiration across European forests during the Anthropocene. *Nat. Clim. Chang.*, 5, 579–583.

Tsilingiris, P.T. (2008). Thermophysical and transport properties of humid air at temperature range between 0 and 100°C. *Energy Convers. Manag.*, 49, 1098–1110.

Ubierna, N. & Farquhar, G.D. (2014). Advances in measurements and models of photosynthetic carbon isotope discrimination in C3 plants. *Plant. Cell Environ.*, 37, 1494–1498.

### Examples

*d13C.to.Ci*(-27, 2015, 900, 24)

---

*d13C.to.CiCa*

*d13C.to.CiCa*

---

### Description

Calculates the ratio of the concentration of leaf intercellular to atmospheric CO<sub>2</sub>, unitless.

### Usage

*d13C.to.CiCa*(*d13C*, year, elevation, temp, frac = 0)

### Arguments

<i>d13C</i>	Measured plant tissue carbon isotope signature, per mille (‰)
year	Year to which the sample corresponds
elevation	Elevation (m.a.s.l.) of the sample, necessary to account for photorespiration processes
temp	Leaf temperature (°C)
frac	Post-photosynthetic fractionation factor, defaults to 0 assuming leaf material, user should supply reasonable value if from wood (generally -1.9 - -2.1)

### Value

The ratio of leaf intercellular to atmospheric CO<sub>2</sub> (Ci/Ca), unitless

## References

- Badeck, F.-W., Tcherkez, G., Nogués, S., Piel, C. & Ghashghaie, J. (2005). Post-photosynthetic fractionation of stable carbon isotopes between plant organs—a widespread phenomenon. *Rapid Commun. Mass Spectrom.*, 19, 1381–1391.
- Belmecheri, S. & Lavergne, A. (2020). Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. *Dendrochronologia*, 63, 125748.
- Bernacchi, C.J., Singaas, E.L., Pimentel, C., Portis Jr, A.R. & Long, S.P. (2001). Improved temperature response functions for models of Rubisco-limited photosynthesis. *Plant, Cell Environ.*, 24, 253–259.
- Craig, H. (1953). The geochemistry of the stable carbon isotopes. *Geochim. Cosmochim. Acta*, 3, 53–92.
- Davies, J.A. & Allen, C.D. (1973). Equilibrium, Potential and Actual Evaporation from Cropped Surfaces in Southern Ontario. *J. Appl. Meteorol.*, 12, 649–657.
- Farquhar, G., O’Leary, M. & Berry, J. (1982). On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.*, 9, 121–137.
- Frank, D.C., Poulter, B., Saurer, M., Esper, J., Huntingford, C., Helle, G., et al. (2015). Water-use efficiency and transpiration across European forests during the Anthropocene. *Nat. Clim. Chang.*, 5, 579–583.
- Tsilingiris, P.T. (2008). Thermophysical and transport properties of humid air at temperature range between 0 and 100°C. *Energy Convers. Manag.*, 49, 1098–1110.
- Ubierna, N. & Farquhar, G.D. (2014). Advances in measurements and models of photosynthetic carbon isotope discrimination in C<sub>3</sub> plants. *Plant. Cell Environ.*, 37, 1494–1498.

## Examples

d13C.to.CiCa(-27, 2015, 900, 24)

---

d13C.to.D13C

*d13C.to.D13C*

---

## Description

Calculates leaf carbon isotope discrimination given plant tissue d13C signature.

## Usage

d13C.to.D13C(d13C, year, frac = 0)

**Arguments**

d13C	Measured plant tissue carbon isotope signature, per mille (‰)
year	Year to which the sample corresponds
frac	Post-photosynthetic fractionation factor, defaults to 0 assuming leaf material, user should supply reasonable value if from wood (generally -1.9 - -2.1)

**Value**

Carbon isotope discrimination in units of per mille (‰)

**References**

- Badeck, F.-W., Tcherkez, G., Nogués, S., Piel, C. & Ghashghaie, J. (2005). Post-photosynthetic fractionation of stable carbon isotopes between plant organs—a widespread phenomenon. *Rapid Commun. Mass Spectrom.*, 19, 1381–1391.
- Belmecheri, S. & Lavergne, A. (2020). Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. *Dendrochronologia*, 63, 125748.
- Craig, H. (1953). The geochemistry of the stable carbon isotopes. *Geochim. Cosmochim. Acta*, 3, 53–92.
- Farquhar, G., O’Leary, M. & Berry, J. (1982). On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.*, 9, 121–137.
- Frank, D.C., Poulter, B., Saurer, M., Esper, J., Huntingford, C., Helle, G., et al. (2015). Water-use efficiency and transpiration across European forests during the Anthropocene. *Nat. Clim. Chang.*, 5, 579–583.
- Ubierna, N. & Farquhar, G.D. (2014). Advances in measurements and models of photosynthetic carbon isotope discrimination in C<sub>3</sub> plants. *Plant. Cell Environ.*, 37, 1494–1498.

**Examples**

d13C.to.D13C(-27, 2015)

---

d13C.to.diffCaCi      *d13C.to.diffCaCi*

---

**Description**

Calculates the difference between the atmospheric CO<sub>2</sub> concentration and the leaf intercellular CO<sub>2</sub> concentration in parts per mil (ppm)

**Usage**

```
d13C.to.diffCaCi(d13C, year, elevation, temp, frac = 0)
```

**Arguments**

d13C	Measured plant tissue carbon isotope signature, per mille (‰)
year	Year to which the sample corresponds
elevation	Elevation (m.a.s.l.) of the sample, necessary to account for photorespiration processes
temp	Leaf temperature (°C)
frac	Post-photosynthetic fractionation factor, defaults to 0 assuming leaf material, user should supply reasonable value if from wood (generally -1.9 - -2.1)

**Value**

The difference between atmospheric and leaf intercellular CO<sub>2</sub> concentrations (ppm).

**References**

- Badeck, F.-W., Tcherkez, G., Nogués, S., Piel, C. & Ghashghaie, J. (2005). Post-photosynthetic fractionation of stable carbon isotopes between plant organs—a widespread phenomenon. *Rapid Commun. Mass Spectrom.*, 19, 1381–1391.
- Belmecheri, S. & Lavergne, A. (2020). Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. *Dendrochronologia*, 63, 125748.
- Bernacchi, C.J., Singaas, E.L., Pimentel, C., Portis Jr, A.R. & Long, S.P. (2001). Improved temperature response functions for models of Rubisco-limited photosynthesis. *Plant, Cell Environ.*, 24, 253–259.
- Craig, H. (1953). The geochemistry of the stable carbon isotopes. *Geochim. Cosmochim. Acta*, 3, 53–92.
- Davies, J.A. & Allen, C.D. (1973). Equilibrium, Potential and Actual Evaporation from Cropped Surfaces in Southern Ontario. *J. Appl. Meteorol.*, 12, 649–657.
- Farquhar, G., O’Leary, M. & Berry, J. (1982). On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.*, 9, 121–137.
- Frank, D.C., Poulter, B., Saurer, M., Esper, J., Huntingford, C., Helle, G., et al. (2015). Water-use efficiency and transpiration across European forests during the Anthropocene. *Nat. Clim. Chang.*, 5, 579–583.
- Tsilingiris, P.T. (2008). Thermophysical and transport properties of humid air at temperature range between 0 and 100°C. *Energy Convers. Manag.*, 49, 1098–1110.
- Ubierna, N. & Farquhar, G.D. (2014). Advances in measurements and models of photosynthetic carbon isotope discrimination in C<sub>3</sub> plants. *Plant. Cell Environ.*, 37, 1494–1498.

**Examples**

```
d13C.to.diffCaCi(-27, 2015, 900, 24)
```

---

```
d13C.to.iWUE
```

```
d13C.to.iWUE
```

---

**Description**

Calculates leaf intrinsic water use efficiency given plant tissue d13C signature.

**Usage**

```
d13C.to.iWUE(d13C, year, elevation, temp, frac = 0)
```

**Arguments**

d13C	Measured plant tissue carbon isotope signature, per mille (‰)
year	Year to which the sample corresponds
elevation	Elevation (m.a.s.l.) of the sample, necessary to account for photorespiration processes
temp	Leaf temperature (°C)
frac	Post-photosynthetic fractionation factor, defaults to 0 assuming leaf material, user should supply reasonable value if from wood (generally -1.9 - -2.1)

**Value**

Intrinsic water use efficiency in units of micromol CO<sub>2</sub> per mol H<sub>2</sub>O

**References**

- Badeck, F.-W., Tcherkez, G., Nogués, S., Piel, C. & Ghashghaie, J. (2005). Post-photosynthetic fractionation of stable carbon isotopes between plant organs—a widespread phenomenon. *Rapid Commun. Mass Spectrom.*, 19, 1381–1391.
- Belmecheri, S. & Lavergne, A. (2020). Compiled records of atmospheric CO<sub>2</sub> concentrations and stable carbon isotopes to reconstruct climate and derive plant ecophysiological indices from tree rings. *Dendrochronologia*, 63, 125748.
- Bernacchi, C.J., Singaas, E.L., Pimentel, C., Portis Jr, A.R. & Long, S.P. (2001). Improved temperature response functions for models of Rubisco-limited photosynthesis. *Plant, Cell Environ.*, 24, 253–259.
- Craig, H. (1953). The geochemistry of the stable carbon isotopes. *Geochim. Cosmochim. Acta*, 3, 53–92.



Davies, J.A. & Allen, C.D. (1973). Equilibrium, Potential and Actual Evaporation from Cropped Surfaces in Southern Ontario. *J. Appl. Meteorol.*, 12, 649–657.

Farquhar, G., O’Leary, M. & Berry, J. (1982). On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.*, 9, 121–137.

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Tsilingiris, P.T. (2008). Thermophysical and transport properties of humid air at temperature range between 0 and 100°C. *Energy Convers. Manag.*, 49, 1098–1110.

Ubierna, N. & Farquhar, G.D. (2014). Advances in measurements and models of photosynthetic carbon isotope discrimination in C3 plants. *Plant. Cell Environ.*, 37, 1494–1498.

### **Examples**

*d13C.to.iWUE*(-27, 2015, 900, 24)

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