

# Package ‘PvSTATEM’

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**Type** Package

**Title** Reading, Quality Control and Preprocessing of MBA (Multiplex Bead Assay) Data

**Description** Speeds up the process of loading raw data from MBA (Multiplex Bead Assay) examinations, performs quality control checks, and automatically normalizes the data, preparing it for more advanced, downstream tasks. The main objective of the package is to create a simple environment for a user, who does not necessarily have experience with R language. The package is developed within the project of the same name - 'PvSTATEM', which is an international project aiming for malaria elimination.

**BugReports** <https://github.com/mini-pw/PvSTATEM/issues>

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create\_standard\_curve\_model\_analyte

*Create a standard curve model for a certain analyte*

---

### Description

Create a standard curve model for a certain analyte

### Usage

```
create_standard_curve_model_analyte(
  plate,
  analyte_name,
  data_type = "Median",
  source_mfi_range_from_all_analytes = FALSE,
  ...
)
```

### Arguments

plate	(Plate()) Object of the Plate class
analyte_name	(character(1)) Name of the analyte for which we want to create the model
data_type	(character(1)) Data type of the value we want to use to fit the model - the same datatype as in the plate file. By default, it equals to Median

source\_mfi\_range\_from\_all\_analytes  
 (logical(1)) If TRUE, the MFI range is calculated from all analytes; if FALSE,  
 the MFI range is calculated only for the current analyte Defaults to FALSE  
 ... Additional arguments passed to the model

**Value**

(Model()) Standard Curve model

---

is\_valid\_data\_type      *Check validity of given data type*

---

**Description**

Check if the data type is valid. The data type is valid if it is one of the elements of the VALID\_DATA\_TYPES vector. The valid data types are:  
 c(Median, Net MFI, Count, Avg Net MFI, Mean, Peak).

**Usage**

```
is_valid_data_type(data_type)
```

**Arguments**

data\_type      A string representing the data type.

**Value**

TRUE if the data type is valid, FALSE otherwise.

---

is\_valid\_sample\_type      *Check validity of given sample type*

---

**Description**

Check if the sample type is valid. The sample type is valid if it is one of the elements of the VALID\_SAMPLE\_TYPES vector. The valid sample types are:  
 c(ALL, BLANK, TEST, NEGATIVE CONTROL, STANDARD CURVE, POSITIVE CONTROL).

**Usage**

```
is_valid_sample_type(sample_type)
```

**Arguments**

sample\_type      A string representing the sample type.

**Value**

TRUE if the sample type is valid, FALSE otherwise.

---

Model	<i>Logistic regression model for the standard curve</i>
-------	---

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**Description**

This model uses the `nplr` package to fit the model. The model is fitted using the formula:

$$y = B + \frac{T - B}{(1 + 10^{b \cdot (x_{mid} - x)})^s},$$

where:

- $y$  is the predicted value, MFI in our case,
- $x$  is the independent variable, dilution in our case,
- $B$  is the bottom plateau - the right horizontal asymptote,
- $T$  is the top plateau - the left horizontal asymptote,
- $b$  is the slope of the curve at the inflection point,
- $x_{mid}$  is the x-coordinate at the inflection point,
- $s$  is the asymmetric coefficient.

This equation is referred to as the Richards' equation. More information about the model can be found in the `nplr` package documentation.

**Public fields**

`analyte` (character(1))  
Name of the analyte for which the model was fitted

`dilutions` (numeric())  
Dilutions used to fit the model

`mfi` (numeric())  
MFI values used to fit the model

`mfi_min` (numeric(1))  
Minimum MFI used for scaling MFI values to the range [0, 1]

`mfi_max` (numeric(1))  
Maximum MFI used for scaling MFI values to the range [0, 1]

`model` (nplr)  
Instance of the `nplr` model fitted to the data

`log_dilution` (logical())  
Indicator should the dilutions be transformed using the `log10` function

`log_mfi` (logical())  
Indicator should the MFI values be transformed using the `log10` function

`scale_mfi` (logical())  
Indicator should the MFI values be scaled to the range [0, 1]

**Active bindings**

top\_asymptote (numeric(1))  
 The top asymptote of the logistic curve

bottom\_asymptote (numeric(1))  
 The bottom asymptote of the logistic curve

**Methods****Public methods:**

- [Model\\$new\(\)](#)
- [Model\\$predict\(\)](#)
- [Model\\$get\\_plot\\_data\(\)](#)
- [Model\\$print\(\)](#)
- [Model\\$clone\(\)](#)

**Method new():** Create a new instance of Model [R6](#) class

*Usage:*

```
Model$new(
  analyte,
  dilutions,
  mfi,
  npars = 5,
  verbose = TRUE,
  log_dilution = TRUE,
  log_mfi = TRUE,
  scale_mfi = TRUE,
  mfi_min = NULL,
  mfi_max = NULL
)
```

*Arguments:*

analyte (character(1))  
 Name of the analyte for which the model was fitted.

dilutions (numeric())  
 Dilutions used to fit the model

mfi MFI (numeric())  
 values used to fit the model

npars (numeric(1))  
 Number of parameters to use in the model

verbose (logical())  
 If TRUE prints messages, TRUE by default

log\_dilution (logical())  
 If TRUE the dilutions are transformed using the log<sub>10</sub> function, TRUE by default

log\_mfi (logical())  
 If TRUE the MFI values are transformed using the log<sub>10</sub> function, TRUE by default

`scale_mfi` (logical())

If TRUE the MFI values are scaled to the range [0, 1], TRUE by default

`mfi_min` (numeric(1))

Enables to set the minimum MFI value used for scaling MFI values to the range [0, 1]. Use values before any transformations (e.g., before the  $\log_{10}$  transformation)

`mfi_max` (numeric(1))

Enables to set the maximum MFI value used for scaling MFI values to the range [0, 1]. Use values before any transformations (e.g., before the  $\log_{10}$  transformation)

**Method** `predict()`: Predict the dilutions from the MFI values

*Usage:*

```
Model$predict(mfi)
```

*Arguments:*

`mfi` (numeric())

MFI values for which we want to predict the dilutions.

*Returns:* (data.frame())

Dataframe with the predicted dilutions, MFI values, and the 97.5% confidence intervals. The columns are named as follows:

- `dilution` - the dilution value
- `dilution.025` - the lower bound of the confidence interval
- `dilution.975` - the upper bound of the confidence interval
- `MFI` - the predicted MFI value

**Method** `get_plot_data()`: Data that can be used to plot the standard curve.

*Usage:*

```
Model$get_plot_data()
```

*Returns:* (data.frame())

Prediction dataframe for scaled MFI (or  $\log_{10}$ MFI) values in the range [0, 1]. Columns are named as in the `predict` method

**Method** `print()`: Function prints the basic information about the model such as the number of parameters or samples used

*Usage:*

```
Model$print()
```

**Method** `clone()`: The objects of this class are cloneable with this method.

*Usage:*

```
Model$clone(deep = FALSE)
```

*Arguments:*

`deep` Whether to make a deep clone.

**Examples**

```

plate_file <- system.file("extdata", "Covid0ISEXPONENT.csv", package = "PvSTATEM")
layout_file <- system.file("extdata", "Covid0ISEXPONENT_layout.csv", package = "PvSTATEM")
plate <- read_luminex_data(plate_file, layout_filepath = layout_file)
model <- create_standard_curve_model_analyte(plate, "S2", log_mfi = TRUE)
print(model)

```

---

Plate	<i>Plate object</i>
-------	---------------------

---

**Description**

A class to represent the luminex plate. It contains information about the samples and analytes that were examined on the plate as well as some additional metadata and batch info

**Public fields**

`plate_name` (character(1))  
Name of the plate.

`analyte_names` (character())  
Names of the analytes that were examined on the plate.

`sample_names` (character())  
Names of the samples that were examined on the plate.

`batch_name` (character(1))  
Name of the batch to which the plate belongs.

`sample_locations` (character())  
Locations of the samples on the plate.

`sample_types` (character())  
Types of the samples that were examined on the plate. The possible values are `c(ALL, BLANK, TEST, NEGATIVE CONTROL, STANDARD CURVE, POSITIVE CONTROL)`.

`dilutions` (character())  
A list containing names of the samples as keys and string representing dilutions as values. The dilutions are represented as strings.

`dilution_values` (numeric())  
A list containing names of the samples as keys and numeric values representing dilutions as values.

`data` (list())  
A list containing dataframes with the data for each sample and analyte. The possible data types - the keys of the list are:  
`c(Median, Net MFI, Count, Avg Net MFI, Mean, Peak)`.  
In each dataframe, the rows represent samples and the columns represent analytes.

`default_data_type` (character(1))  
The default data type that will be returned by the `get_data` method. By default is set to Median.

`batch_info (list())`

A list containing additional, technical information about the batch.

`layout (character())`

A list containing information about the layout of the plate. The layout is read from the separate file and usually provides additional information about the dilutions, sample names, and the sample layout on the actual plate.

`blank_adjusted (logical)`

A flag indicating whether the blank values have been adjusted.

## Methods

### Public methods:

- [Plate\\$new\(\)](#)
- [Plate\\$print\(\)](#)
- [Plate\\$summary\(\)](#)
- [Plate\\$get\\_data\(\)](#)
- [Plate\\$get\\_dilution\(\)](#)
- [Plate\\$get\\_dilution\\_values\(\)](#)
- [Plate\\$blank\\_adjustment\(\)](#)
- [Plate\\$clone\(\)](#)

**Method** `new()`: Method to initialize the Plate object

*Usage:*

```
Plate$new(
  plate_name,
  sample_names,
  analyte_names,
  batch_name = "",
  dilutions = NULL,
  dilution_values = NULL,
  sample_types = NULL,
  data = NULL,
  sample_locations = NULL,
  default_data_type = NULL,
  batch_info = NULL,
  layout = NULL
)
```

*Arguments:*

`plate_name (character(1))`

Name of the plate. By default is set to an empty string, during the reading process it is set to the name of the file from which the plate was read.

`sample_names (character())`

Names of the samples that were examined on the plate.

`analyte_names (character())`

Names of the analytes that were examined on the plate.



`batch_name` (character(1))  
 Name of the batch to which the plate belongs. By default is set to an empty string, during the reading process it is set to the batch field of the plate

`dilutions` (character())  
 A list containing names of the samples as keys and string representing dilutions as values. The dilutions are represented as strings.

`dilution_values` (numeric())  
 A list containing names of the samples as keys and numeric values representing dilutions as values.

`sample_types` (character())  
 Types of the samples that were examined on the plate. The possible values are c(ALL, BLANK, TEST, NEGATIVE CONTROL, STANDARD CURVE, POSITIVE CONTROL).

`data` (list())  
 A list containing dataframes with the data for each sample and analyte. The possible data types - the keys of the list are c(Median, Net MFI, Count, Avg Net MFI, Mean, Peak). In each dataframe, the rows represent samples and the columns represent analytes.

`sample_locations` (character())  
 Locations of the samples on the plate.

`default_data_type` (character(1))  
 The default data type that will be returned by the `get_data` method. By default is set to Median.

`batch_info` (list())  
 A list containing additional, technical information about the batch.

`layout` (character())  
 A list containing information about the layout of the plate. The layout is read from the separate file and usually provides additional information about the dilutions, sample names, and the sample layout on the actual plate.

**Method** `print()`: Function prints the basic information about the plate such as the number of samples and analytes

*Usage:*

```
Plate$print(...)
```

*Arguments:*

... Additional parameters to be passed to the print function Print the summary of the plate

**Method** `summary()`: Function outputs basic information about the plate, such as examination date, batch name, and sample types.

*Usage:*

```
Plate$summary(..., include_names = FALSE)
```

*Arguments:*

... Additional parameters to be passed to the print function Get data for a specific analyte and sample type

`include_names` If `include_names` parameter is TRUE, a part from count of control samples, provides also their names. By default FALSE

**Method** `get_data()`: Function returns data for a specific analyte and sample.

*Usage:*

```
Plate$get_data(
  analyte,
  sample_type = "ALL",
  data_type = self$default_data_type
)
```

*Arguments:*

`analyte` An analyte name or its id of which data we want to extract. If set to 'ALL' returns data for all analytes.

`sample_type` is a type of the sample we want to extract data from. The possible values are `c(ALL, BLANK, TEST, NEGATIVE CONTROL, STANDARD CURVE, POSITIVE CONTROL)`. Default value is ALL.

`data_type` The parameter specifying which data type should be returned. This parameter has to take one of values:

`c(Median, Net MFI, Count, Avg Net MFI, Mean, Peak)`. What's more, the `data_type` has to be present in the plate's data Default value is `plate$default_data_type`, which is usually Median.

*Returns:* Dataframe containing information about a given sample type and analyte Get the string representation of dilutions

**Method** `get_dilution()`: Function returns the dilution represented as strings for a specific sample type.

*Usage:*

```
Plate$get_dilution(sample_type)
```

*Arguments:*

`sample_type` type of the samples that we want to obtain the dilution for. The possible values are

`c(ALL, BLANK, TEST, NEGATIVE CONTROL, STANDARD CURVE, POSITIVE CONTROL)` Default value is ALL.

*Returns:* A list containing names of the samples as keys and string representing dilutions as values. Get the numeric representation of dilutions

**Method** `get_dilution_values()`: Function returns the dilution values for a specific sample type.

*Usage:*

```
Plate$get_dilution_values(sample_type)
```

*Arguments:*

`sample_type` type of the samples that we want to obtain the dilution values for. The possible values are

`c(ALL, BLANK, TEST, NEGATIVE CONTROL, STANDARD CURVE, POSITIVE CONTROL)` Default value is ALL.

*Returns:* A list containing names of the samples as keys and numeric values representing dilutions as values.

Adjust the MFI values by subtracting the background

**Method** `blank_adjustment()`: Function adjusts the values of samples (all samples excluding the blanks) by clamping the values to the aggregated value of the BLANK samples for each analyte separately.

The purpose of this operation is to unify the data by clamping values below the background noise. how this method works was inspired by the paper <https://doi.org/10.1038/s41598-020-57876-0> which covers the quality control in the MBA.

In short, this operation firstly calculates the aggregate of MFI in the BLANK samples (available methods are: min, max, mean, median) and then replaces all values below this threshold with the threshold value.

Method does not modifies the data of type Count.

This operation is recommended to be performed before any further analysis, but is optional. Skipping it before further analysis is allowed, but will result in a warning.

*Usage:*

```
Plate$blank_adjustment(threshold = "max", in_place = TRUE)
```

*Arguments:*

`threshold` The method used to calculate the background value for each analyte. Every value below this threshold will be clamped to the threshold value. By default max. Available methods are: min, max, mean, median.

`inplace` Whether the method should produce new plate with adjusted values or not, By default TRUE - operates on the current plate.

**Method** `clone()`: The objects of this class are cloneable with this method.

*Usage:*

```
Plate$clone(deep = FALSE)
```

*Arguments:*

`deep` Whether to make a deep clone.

---

PlateBuilder

*PlateBuilder*

---

## Description

This class helps creating the Plate object. It is used to store the data and validate the final fields.

## Active bindings

`layout_as_vector` Print the layout associated with the plate as a flattened vector of values.

## Methods

### Public methods:

- [PlateBuilder\\$new\(\)](#)
- [PlateBuilder\\$set\\_sample\\_locations\(\)](#)
- [PlateBuilder\\$set\\_dilutions\(\)](#)

- `PlateBuilder$set_sample_types()`
- `PlateBuilder$set_sample_names()`
- `PlateBuilder$set_data()`
- `PlateBuilder$set_default_data_type()`
- `PlateBuilder$set_batch_info()`
- `PlateBuilder$set_plate_name()`
- `PlateBuilder$set_layout()`
- `PlateBuilder$build()`
- `PlateBuilder$clone()`

**Method** `new()`: Initialize the PlateBuilder object

*Usage:*

```
PlateBuilder$new(sample_names, analyte_names, batch_name = "", verbose = TRUE)
```

*Arguments:*

`sample_names` • vector of sample names measured during an examination in the same order as in the data

`analyte_names` • vector of analytes names measured during an examination in the same order as in the data

`batch_name` • name of the batch during which the plate was examined obtained from the plate info. An optional parameter, by default set to "" - an empty string.

`verbose` • logical value indicating whether to print additional information. This parameter is stored as a private attribute of the object and reused in other methods

**Method** `set_sample_locations()`: Set the sample types used during the examination

*Usage:*

```
PlateBuilder$set_sample_locations(sample_locations)
```

*Arguments:*

`sample_locations` vector of sample locations pretty name ie. A1, B2

**Method** `set_dilutions()`: Extract and set the dilutions from layout, sample names or use a provided vector of values. The provided vector should be the same length as the number of samples and should contain dilution factors saved as strings

*Usage:*

```
PlateBuilder$set_dilutions(use_layout_dilutions = TRUE, values = NULL)
```

*Arguments:*

`use_layout_dilutions` logical value indicating whether to use names extracted from layout files to extract dilutions. If set to FALSE the function uses the sample names as a source for dilution

`values` a vector of dilutions to overwrite the extraction process

Set and extract sample types from the sample names. Optionally use the layout file to extract the sample types

**Method** `set_sample_types()`:

*Usage:*

```
PlateBuilder$set_sample_types(use_layout_types = TRUE, values = NULL)
```

*Arguments:*

`use_layout_types` logical value indicating whether to use names extracted from layout files to extract sample types

`values` a vector of sample types to overwrite the extraction process

**Method** `set_sample_names()`: Set the sample names used during the examination. If the layout is provided, extract the sample names from the layout file. Otherwise, uses the original sample names from the Luminex file

*Usage:*

```
PlateBuilder$set_sample_names(use_layout_sample_names = TRUE)
```

*Arguments:*

`use_layout_sample_names` logical value indicating whether to use names extracted from layout files. If set to false, this function only checks if the sample names are provided in the plate

**Method** `set_data()`: Set the data used during the examination

*Usage:*

```
PlateBuilder$set_data(data)
```

*Arguments:*

`data` a named list of data frames containing information about the samples and analytes. The list is named by the type of the data e.g. Median, Net MFI, etc. The data frames contain information about the samples and analytes. The rows are different measures, whereas the columns represent different analytes. Example of how `data$Median` looks like:

Sample	Analyte1	Analyte2	Analyte3
Sample1	1.2	2.3	3.4
Sample2	4.5	5.6	6.7
...	...	...	...
Sample96	7.8	8.9	9.0

**Method** `set_default_data_type()`: Set the data type used for calculations

*Usage:*

```
PlateBuilder$set_default_data_type(data_type = "Median")
```

*Arguments:*

`data_type` a character value representing the type of data that is currently used for calculations. By default, it is set to Median

**Method** `set_batch_info()`: Set the batch info for the plate

*Usage:*

```
PlateBuilder$set_batch_info(batch_info)
```

*Arguments:*

`batch_info` a raw list containing metadata about the plate read from the Luminex file

**Method** `set_plate_name()`: Set the plate name for the plate. The plate name is extracted from the filepath

*Usage:*

```
PlateBuilder$set_plate_name(file_path)
```

**Method** `set_layout()`: Set the layout matrix for the plate. This function performs basic validation

- verifies if the plate is a matrix of shape 8x12 with 96 wells

*Usage:*

```
PlateBuilder$set_layout(layout_matrix)
```

*Arguments:*

`layout_matrix` a matrix containing information about the sample names, dilutions, etc.

**Method** `build()`: Create a Plate object from the PlateBuilder object

*Usage:*

```
PlateBuilder$build(validate = TRUE)
```

**Method** `clone()`: The objects of this class are cloneable with this method.

*Usage:*

```
PlateBuilder$clone(deep = FALSE)
```

*Arguments:*

`deep` Whether to make a deep clone.

plot\_counts

*Plot counts in a 96-well plate*

## Description

This is a function used to plot counts in a 96-well plate using a color to represent the count ranges. There is possibility to plot exact counts in each well.

If plot window is resized, it's best to re-run the function to adjust the scaling. Sometimes when legend is plotted, whole layout may be shifted, then it's best to stretch the window, and everything will be adjusted automatically.

## Usage

```
plot_counts(
  plate,
  analyte_name,
  plot_counts = TRUE,
  plot_legend = FALSE,
  lower_threshold = 50,
  higher_threshold = 70
)
```

**Arguments**

plate	The plate object with the counts data
analyte_name	The name of the analyte
plot_counts	Logical indicating if the counts should be plotted
plot_legend	Logical indicating if the legend should be plotted
lower_threshold	The lower threshold for the counts, it separates green and yellow colors
higher_threshold	The higher threshold for the counts, it separates yellow and red colors

**Value**

A ggplot object

**Examples**

```

plate_filepath <- system.file("extdata", "CovidOISEXPONENT_CO.csv",
  package = "PvSTATEM", mustWork = TRUE
)
layout_filepath <- system.file("extdata", "CovidOISEXPONENT_CO_layout.xlsx",
  package = "PvSTATEM", mustWork = TRUE
)
plate <- read_luminex_data(plate_filepath, layout_filepath)
plot_counts(
  plate = plate, analyte_name = "OC43_NP_NA",
  plot_counts = TRUE, plot_legend = FALSE
)

```

---

plot_layout	<i>Plot layout of a 96-well plate</i>
-------------	---------------------------------------

---

**Description**

This is a function used to plot the layout of a 96-well plate using a color to represent the sample types.

If plot window is resized, it's best to re-run the function to adjust the scaling. Sometimes when legend is plotted, whole layout may be shifted, then it's best to stretch the window, and everything will be adjusted automatically.

**Usage**

```
plot_layout(plate, plot_legend = TRUE)
```

**Arguments**

plate            The plate object with the layout information  
plot\_legend     Logical indicating if the legend should be plotted

**Value**

A ggplot object

**Examples**

```
plate_filepath <- system.file("extdata", "CovidOISEXPONENT_CO.csv",  
  package = "PvSTATEM", mustWork = TRUE  
)  
layout_filepath <- system.file("extdata", "CovidOISEXPONENT_CO_layout.xlsx",  
  package = "PvSTATEM", mustWork = TRUE  
)  
plate <- read_luminex_data(plate_filepath, layout_filepath)  
plot_layout(plate = plate, plot_legend = TRUE)
```

---

plot\_mfi\_for\_analyte    *Plot MFI value distribution for a given analyte*

---

**Description**

Plot MFI value distribution for a given analyte

**Usage**

```
plot_mfi_for_analyte(  
  plate,  
  analyte_name,  
  data_type = "Median",  
  plot_type = "boxplot"  
)
```

**Arguments**

plate            A plate object  
analyte\_name     The analyte to plot  
data\_type        The type of data to plot. Default is "Median"  
plot\_type        The type of plot to generate. Default is "boxplot". Available options are "box-plot" and "violin".

**Value**

A ggplot object



---

plot\_standard\_curve\_analyte  
*Standard curves*

---

### Description

Plot standard curve samples of a plate of a given analyte.

### Usage

```
plot_standard_curve_analyte(  
  plate,  
  analyte_name,  
  data_type = "Median",  
  decreasing_dilution_order = TRUE,  
  log_scale = c("all"),  
  plot_line = TRUE,  
  plot_blank_mean = TRUE,  
  plot_dilution_bounds = TRUE,  
  verbose = TRUE  
)
```

### Arguments

plate	A plate object
analyte_name	Name of the analyte of which standard curve we want to plot.
data_type	Data type of the value we want to plot - the same datatype as in the plate file. By default equals to Net MFI
decreasing_dilution_order	If TRUE the dilutions are plotted in decreasing order, TRUE by default
log_scale	Which elements on the plot should be displayed in log scale. By default "dilutions". If NULL or c() no log scale is used, if "all" or c("dilutions", "MFI") all elements are displayed in log scale.
plot_line	If TRUE a line is plotted, TRUE by default
plot_blank_mean	If TRUE the mean of the blank samples is plotted, TRUE by default
plot_dilution_bounds	If TRUE the dilution bounds are plotted, TRUE by default
verbose	If TRUE prints messages, TRUE by default

### Value

ggplot object with the plot

---

```
plot_standard_curve_analyte_with_model
```

*Plot standard curve of a certain analyte with fitted model*

---

### Description

Function plots the values of standard curve samples and the fitted model.

### Usage

```
plot_standard_curve_analyte_with_model(  
  plate,  
  model,  
  data_type = "Median",  
  decreasing_dilution_order = TRUE,  
  log_scale = c("all"),  
  plot_asymptote = TRUE,  
  plot_test_predictions = TRUE,  
  plot_blank_mean = TRUE,  
  plot_dilution_bounds = TRUE,  
  verbose = TRUE  
)
```

### Arguments

plate	Plate object
model	fitted Model object, which predictions we want to plot
data_type	Data type of the value we want to plot - the same datatype as in the plate file. By default equals to Median
decreasing_dilution_order	If TRUE the dilutions are plotted in decreasing order, TRUE by default.
log_scale	Which elements on the plot should be displayed in log scale. By default "all". If NULL or c() no log scale is used, if "all" or c("dilutions", "MFI") all elements are displayed in log scale.
plot_asymptote	If TRUE the asymptotes are plotted, TRUE by default
plot_test_predictions	If TRUE the predictions for the test samples are plotted, TRUE by default The predictions are obtained through extrapolation of the model
plot_blank_mean	If TRUE the mean of the blank samples is plotted, TRUE by default
plot_dilution_bounds	If TRUE the dilution bounds are plotted, TRUE by default
verbose	If TRUE prints messages, TRUE by default

**Value**

a ggplot object with the plot

---

predict.Model	<i>Predict the dilutions from the MFI values</i>
---------------	--

---

**Description**

More details can be found here: [Model](#)

**Usage**

```
## S3 method for class 'Model'
predict(object, mfi, ...)
```

**Arguments**

object	(Model()) Object of the Model class
mfi	(numeric()) MFI values for which we want to predict the dilutions. Should be in the same scale as the MFI values used to fit the model
...	Additional arguments passed to the method

**Value**

(data.frame())

---

process_plate	<i>Process a plate and save computed dilutions to a CSV</i>
---------------	---

---

**Description**

The behavior can be summarized as follows:

1. Adjust blanks if not already done.
2. Fit a model to each analyte using standard curve samples.
3. Compute dilutions for each analyte using the corresponding model.
4. Aggregate computed dilutions into a single data frame.
5. Save the computed dilutions to a CSV file.

**Usage**

```
process_plate(
  plate,
  output_path = NULL,
  data_type = "Median",
  adjust_blanks = FALSE,
  verbose = TRUE,
  ...
)
```

**Arguments**

plate	(Plate()) a plate object
output_path	(character(1)) path to save the computed dilutions. If not provided the file will be saved in the working directory with the name dilutions_{plate_name}.csv. Where the {plate_name} is the name of the plate.
data_type	(character(1)) type of data to use for the computation. Median is the default
adjust_blanks	(logical(1)) adjust blanks before computing dilutions. Default is FALSE
verbose	(logical(1)) print additional information. Default is TRUE
...	Additional arguments to be passed to the fit model function (create_standard_curve_model_analyte)

**Examples**

```
plate_file <- system.file("extdata", "CovidOISEXPONENT.csv", package = "PvSTATEM")
layout_file <- system.file("extdata", "CovidOISEXPONENT_layout.csv", package = "PvSTATEM")

plate <- read_luminex_data(plate_file, layout_file)

tmp_dir <- tempdir(check = TRUE)
temporary_filepath <- file.path(tmp_dir, "output.csv")
process_plate(plate, output_path = temporary_filepath)
# create and save dataframe with computed dilutions
```

---

```
read_intelliflex_format
```

*Read the Intelliflex format data*

---

**Description**

Read the Intelliflex format data

**Usage**

```
read_intelliflex_format(path, verbose = TRUE)
```

**Arguments**

path	Path to the INTELLIFLEX file
verbose	Print additional information. Default is TRUE

---

read_layout_data	<i>Read layout data from a file</i>
------------------	-------------------------------------

---

**Description**

Read layout data from a file

**Usage**

```
read_layout_data(layout_file_path, ...)
```

**Arguments**

layout_file_path	Path to the layout file
...	Additional arguments to pass to the underlying read function

**Value**

A matrix with the layout data. The row names are supposed to be letters A,B,C, etc. The column names are supposed to be numbers 1,2,3, etc.

---

read_luminex_data	<i>Read Luminex Data</i>
-------------------	--------------------------

---

**Description**

Reads a file containing Luminex data and returns a Plate object. If provided, can also read a layout file, which usually contains information about the sample names, sample types or its dilutions.

The function is capable of reading data in two different formats:

- xPONENT
- INTELLIFLEX which are produced by two different Luminex machines.

**Usage**

```
read_luminex_data(  
  plate_filepath,  
  layout_filepath = NULL,  
  format = "xPONENT",  
  plate_file_separator = ",",  
  plate_file_encoding = "UTF-8",  
  use_layout_sample_names = TRUE,  
  use_layout_types = TRUE,  
  use_layout_dilutions = TRUE,  
  default_data_type = "Median",  
  sample_types = NULL,  
  dilutions = NULL,  
  verbose = TRUE  
)
```

**Arguments**

`plate_filepath` Path to the Luminex plate file

`layout_filepath`  
Path to the Luminex layout file

`format` The format of the Luminex data. Select from: xPONENT, INTELLIFLEX

`plate_file_separator`  
The separator used in the plate file

`plate_file_encoding`  
The encoding used in the plate file

`use_layout_sample_names`  
Whether to use names from the layout file in extracting sample names.

`use_layout_types`  
Whether to use names from the layout file in extracting sample types. Works only when layout file is provided

`use_layout_dilutions`  
Whether to use dilutions from the layout file in extracting dilutions. Works only when layout file is provided

`default_data_type`  
The default data type to use if none is specified

`sample_types` a vector of sample types to use instead of the extracted ones

`dilutions` a vector of dilutions to use instead of the extracted ones

`verbose` Whether to print additional information and warnings. TRUE by default

**Value**

Plate file containing the Luminex data

## Examples

```
plate_file <- system.file("extdata", "Covid0ISEXPONENT.csv", package = "PvSTATEM")
layout_file <- system.file("extdata", "Covid0ISEXPONENT_layout.csv", package = "PvSTATEM")
plate <- read_luminex_data(plate_file, layout_file)

plate_file <- system.file("extdata", "Covid0ISEXPONENT_CO.csv", package = "PvSTATEM")
layout_file <- system.file("extdata", "Covid0ISEXPONENT_CO_layout.xlsx", package = "PvSTATEM")
# To suppress warnings and additional information use verbose = FALSE
plate <- read_luminex_data(plate_file, layout_file, verbose = FALSE)
```

---

read\_xponent\_format    *Read the xPONENT format data*

---

## Description

Read the xPONENT format data

## Usage

```
read_xponent_format(
  path,
  exact_parse = FALSE,
  encoding = "utf-8",
  separator = ",",
  verbose = TRUE
)
```

## Arguments

path	Path to the xPONENT file
exact_parse	Whether to parse the file exactly or not Exact parsing means that the batch, calibration and assay metadata will be parsed as well
encoding	Encoding of the file
separator	Separator for the CSV values
verbose	Whether to print the progress. Default is TRUE

---

```
translate_sample_names_to_sample_types
```

*Translate sample names to sample types*

---

## Description

Function translates sample names to sample types based on the sample name from Luminex file and the sample name from the layout file, which may not be provided. The function uses regular expressions to match the sample names to the sample types.

It parses the names as follows:

If `sample_names` or `sample_names_from_layout` equals to BLANK, BACKGROUND or B, then `SampleType` equals to BLANK

If `sample_names` or `sample_names_from_layout` equals to STANDARD CURVE, SC, S, contains substring `1/\d+` and has prefix `, S_, S , S` or CP3, then `SampleType` equals to STANDARD CURVE

If `sample_names` or `sample_names_from_layout` equals to NEGATIVE CONTROL, N, or contains substring NEG, then `SampleType` equals to NEGATIVE CONTROL

If `sample_names` or `sample_names_from_layout` starts with P followed by whitespace, POS followed by whitespace, some sample name followed by substring `1/\d+` `SampleType` equals to POSITIVE CONTROL

Otherwise, the returned `SampleType` is TEST

## Usage

```
translate_sample_names_to_sample_types(  
  sample_names,  
  sample_names_from_layout = NULL  
)
```

## Arguments

`sample_names` (character())  
Vector of sample names from Luminex file

`sample_names_from_layout`  
(character())  
Vector of sample names from Layout file values in this vector may be different than `sample_names` and may contain additional information about the sample type like dilution. This vector when set has to have at least the length of `sample_names`.

## Value

A vector of valid `sample_type` strings of length equal to the length of `sample_names`



**Examples**

```
translate_sample_names_to_sample_types(c("B", "BLANK", "NEG", "TEST1"))  
translate_sample_names_to_sample_types(c("S", "CP3"))
```

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